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Photoswitching of red cyanine dyes

Fluorescent **650 nm**
+ thiol

Cy5 / Alexa 647

photoactivation ↑
Deactivation ↓

360 nm Dark
650 nm

Fluorescent **Dark**

RS⁻ hu

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B. Huang (2009); Dempsey et al. (2009)

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Switchable probes commercially available

400 500 600 700 nm

Cyanine dye + thiol system
Alexa647 Cy5 Cy7
Bates et al., 2005, Bates et al., 2007, Huang et al., 2008

Rhodamine dye + redox system
Alexa488 Atto565 Alexa532 Atto590 Atto520 Alexa568 Atto655 Atto700
Heilemann et al., 2009

Photoactivatable fluorescent proteins
PA-GFP mEosFP2 PS-CFP2 Dendra2 Dronpa EYFP PanCherry
Reviews: Lukyanov et al., Nat. Rev. Cell Biol., 2005
Lippincott-Schwartz et al., Trends Cell Biol., 2009

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B. Huang (2009)

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PALM: Photoactivated Localization Microscopy

ACTIVATED BLEACHED ACTIVATED BLEACHED

Betzig et al., Science (2006)

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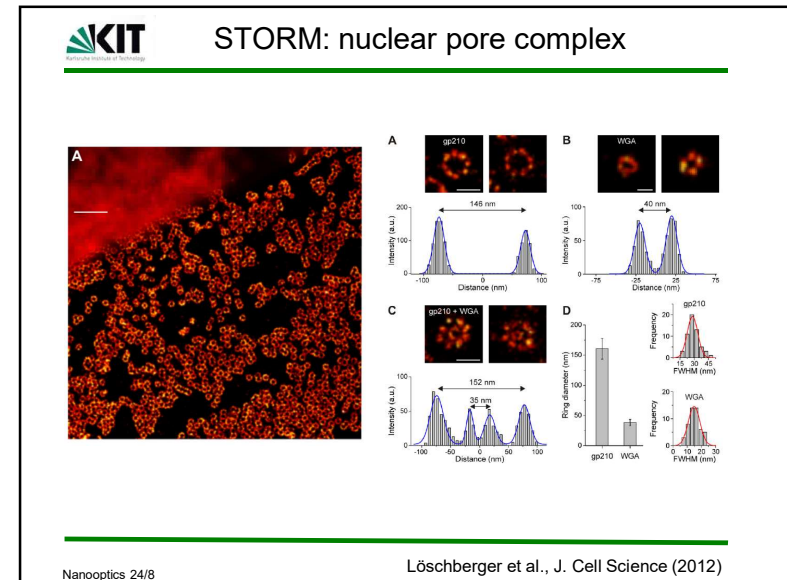
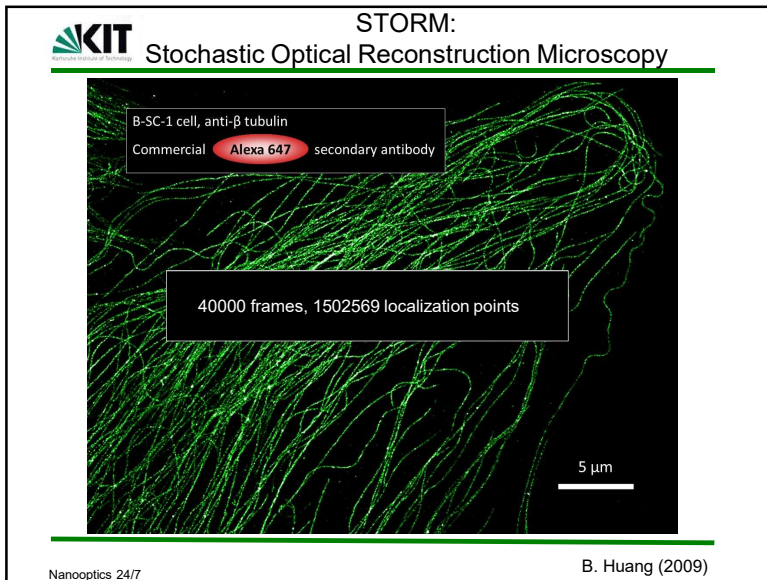
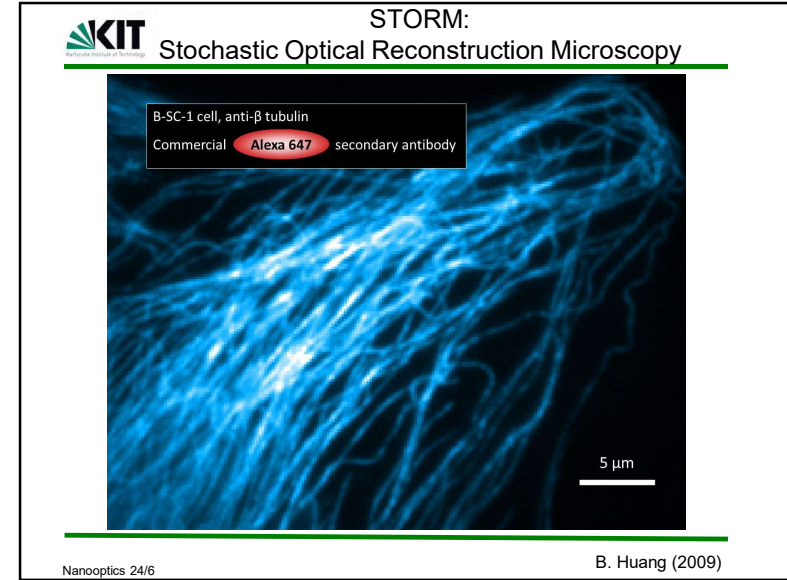
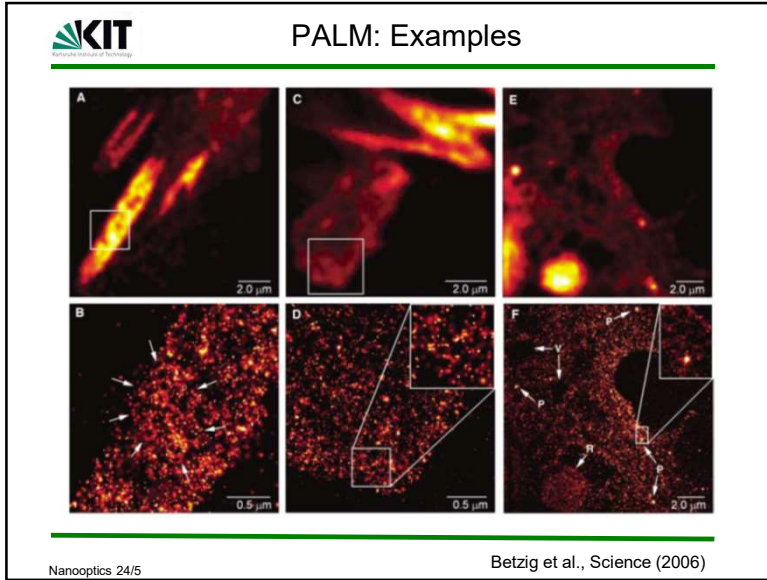
PALM: Examples

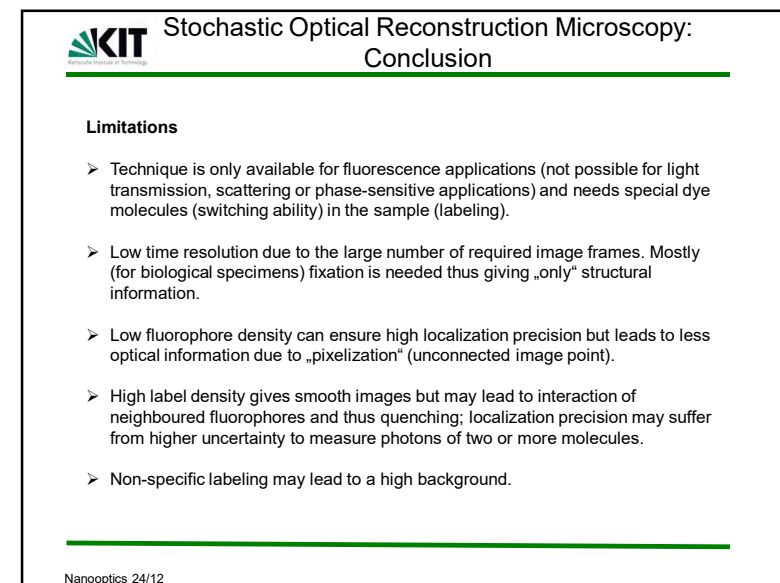
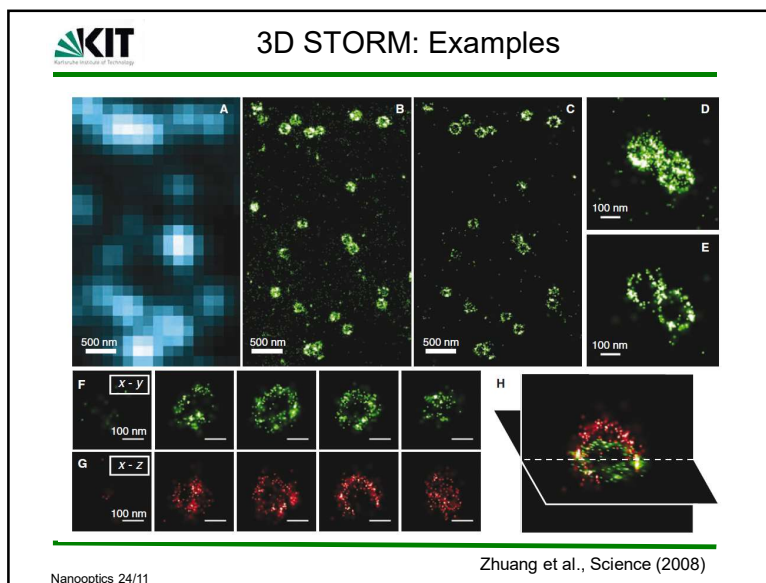
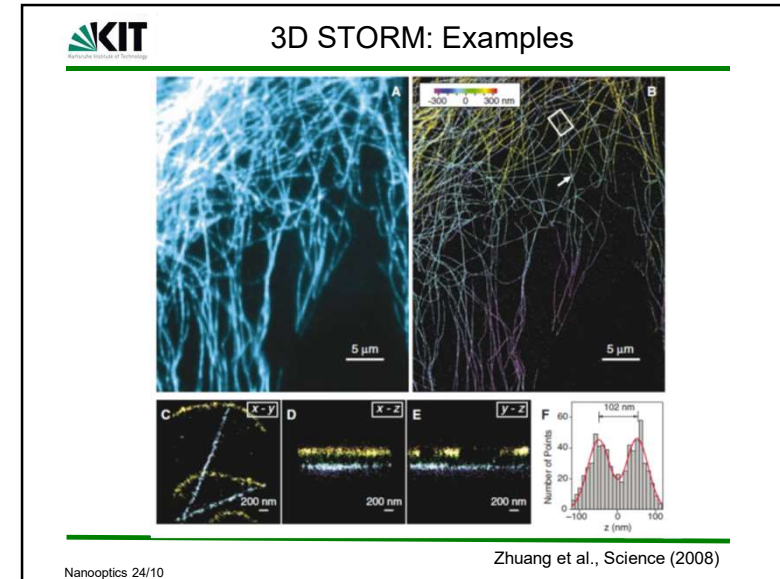
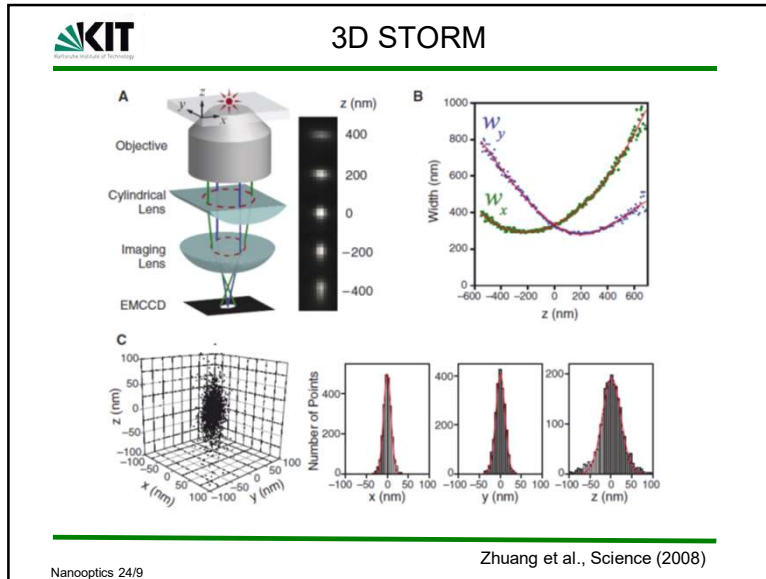
A B

1.0 μm 1.0 μm

Betzig et al., Science (2006)

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4. Nano-optics using far-field optical techniques

- 4.1 Introduction: single-molecule methods in biology
- 4.2 Single-molecule tracking (SMT)
- 4.3 Stochastic optical reconstruction microscopy (STORM)
- 4.4 4pi microscopy
- 4.5 Stimulated emission depletion (STED)
- 4.6 3D laser lithography using STED

4Pi Microscopy

The task:

- Increase resolution in fluorescence microscopy over classical Abbé Limits

$$FWHM_{xy} = \frac{0.4\lambda}{n \cdot \sin \alpha}$$

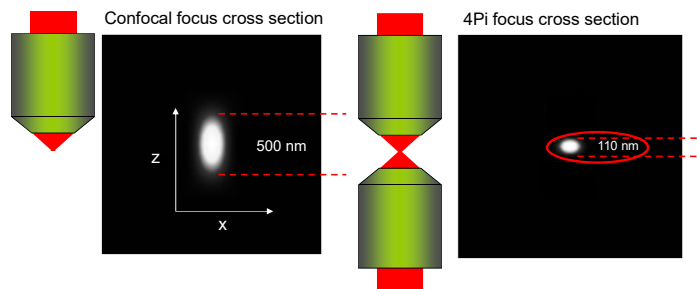
$$FWHM_z = \frac{0.45\lambda}{n(1 - \cos \alpha)}$$

- xy resolution: ~ 200nm

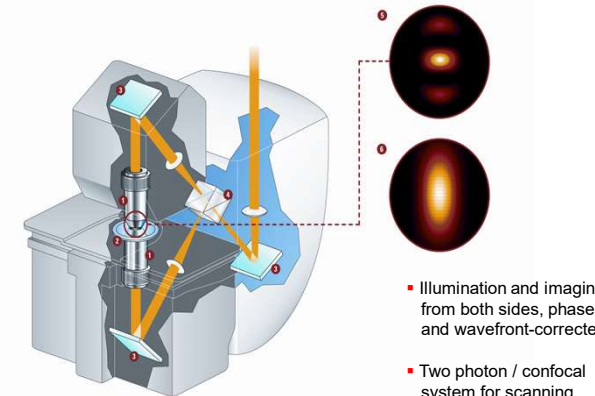
- z resolution (confocal): ~ 500nm

4Pi Microscopy

- 4Pi microscopy combines the Numerical Aperture of 2 opposing objectives through interference
- The 4Pi focus volume is a factor 3 – 5 times smaller than the confocal focus
- The axial resolution at the excitation wavelength of 780 nm is 110nm
- Interference side lobes are removed by linear point deconvolution



Central element: the 4Pi Interferometer



- Illumination and imaging from both sides, phase- and wavefront-corrected

- Two photon / confocal system for scanning

KIT **4Pi Interferometer controls**

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KIT **Objectives for 4Pi - outstanding specifications**

Specifications:

- Focus aberration of each individual objective: max. 300nm in the range 488nm to 633nm
- Objective pairs must be identical within +/- 50 nm in the range 488nm - 633nm.
- Color magnification error must be less than 1% within the range of 488nm - 633nm.
- Magnification of both objectives must be identical within 0,1%

→ Matching objective pairs need to be selected

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KIT **4Pi Microscopy: Sample preparation**

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KIT **Performance at theoretical limits**

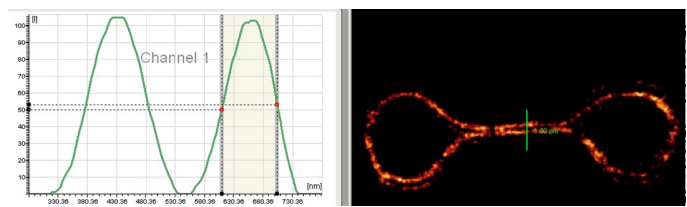
- Calculated (blue) and measured (red) optical resolution for a thin structure (mirror)

Wavelength: 840nm
Objective: 1.4 NA (Oil Immersion)

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Parasitology: Human red blood cell - membrane separation by 4Pi microscopy



Channel 1

- Z resolution ~ 100nm
- Clear separation of membranes 100 nm apart
- Cell membrane (Band-III-protein) labelled with Qdot@ 585-coupled antibodies

Dr. J. A. Dvorak and Dr. F. Tokumasu, National Institute of Allergy and Infectious Diseases, NIH, Washington (USA)

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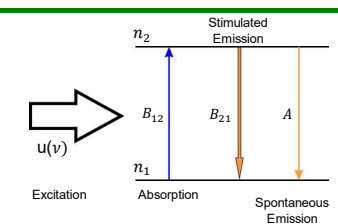
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Stimulated Emission



Excitation $u(\nu)$

Absorption B_{12}

Spontaneous Emission A

Stimulated Emission B_{21}

$B_{12} = B_{21} = B$ for non-degenerate states

$$\dot{n}_2 = B u(\nu) n_1 - B u(\nu) n_2 - A n_2$$

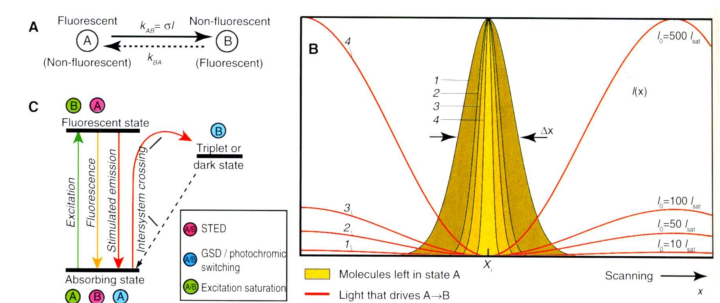
The coefficient **A** (spontaneous emission) depends on the density of states:

$$\frac{A}{8\pi \nu^2/c^3} = B \cdot h\nu$$

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STED: Confinement by "Inverse Saturation" or "Saturated Depletion"



A Fluorescent (A) ↔ Non-fluorescent (B) with $k_{AB} = \sigma I$ and k_{BA}

B Intensity profile $I(x)$ with $I_0 = 500 I_{sat}$ and $I_0 = 10 I_{sat}$

C Energy level diagram showing transitions: Excitation, Fluorescence, Stimulated emission, Intersystem crossing to Triplet or dark state, and GSD / photochromic switching.

Legend:

- STED (red arrow)
- GSD / photochromic switching (blue arrow)
- Excitation saturation (green arrow)

Yellow area: Molecules left in state A

Red line: Light that drives A → B

Hell et al., in Pawley (ed.), Biological Confocal Microscopy (2006)

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